An Advantageous Synthesis of 1D- and 1L-1,2,3,5/4-Cyclohexanepentol

by Marco A. Biamonte and Andrea Vasella*

Laboratorium für Organische Chemie, ETH-Zentrum, Universitätstrasse 16, CH-8092 Zürich

The title compounds D-10 and L-10 were prepared from 1 in eight steps and in a combined overall yield of 41-49%.

Introduction. – myo-Inositol (1) analogues have attracted a great deal of attention due to the important biological role of the inositol phosphates [1] and of the glycosylphosphatidylinositol (GPI) anchors [2]. However, relatively few modifications of the enantiotopic HO–C(4) and HO–C(6) groups of D-myo-inositol have been reported ¹). In the context of our studies on intramolecular H-bonding [12], we required orthoester derivatives of 1D-1,2,3,5/4-cyclohexanepentol (D-10) and 1L-1,2,3,5/4-cyclohexanepentol (L-10) ('6-deoxy-D-myo-inositol' and '4-deoxy-D-myo-inositol', resp.). A synthesis of D-10 from methyl β -D-galactopyranoside has been already described (seven steps, 11% overall yield) [6]. However, to the best of our knowledge, no synthesis of L-10 has been reported, and the known preparations of DL-10 are not satisfactory²). The orthoformates D-5 and L-5 have been prepared from myo-inositol in seven steps, and in a combined overall yield of 9%, but have not been deprotected [13]. We now report an optimized synthesis of their silylated orthopentanoate analogues D-11 and L-11 using a similar strategy, and describe their transformation into D-10 and L-10.

Results and Discussion. – myo-Inositol (1; Scheme) was treated with $BuC(OMe)_3$ and a small amount of camphorsulfonic acid to give the orthoester 2 (91%). The advantages associated with the formation of the orthopentanoate 2, as compared to the orthoformate 3 [14], are due to the lower temperature required for the orthoesterification and to the much easier purification of 2. This is both due to the cleaner reaction (no benzoquinone is formed, as in the synthesis of 3) and to the much higher solubility. Similarly as for 3 [15], HO-C(2) acts as H-bond donor in a bifurcated, intramolecular H-bond, and the HO-C(4) and HO-C(6) groups form a strong intramolecular H-bond, with one of them acting as donor, and the other as acceptor. This leads to markedly different nucleophilicities for the three OH groups [15]. This property was exploited for the selective monothiocarbonylation of 2 by 4-O-tolyl thiochloroformate. The regioselectivity of this acylation is evident from the lack of symmetry expressed in the ¹H-NMR spectrum of the resulting carbonothioate DL-4 that was isolated in 91% by chromatog-

¹) These OH groups have been replaced by H [3-6], NH₂ [7], MeO [8], and F [4][5][9] but not, to our knowledge, by SH, Cl, Br, or I. For the biological activity of these analogues, see [7][10][11].

²) The pentol DL-10 was obtained from myo-inositol in five steps (1% overall yield) [4], or in seven steps (ca. 1% overall yield) [3], and from cis-cyclohexa-3,5-diene-1,2-diol in ten steps (15% overall yield) [5].

raphy or in 61% by direct crystallization. Barton-McCombie deoxygenation of DL-4 with Bu₃SnH gave the diol DL-6 (80–93%). The new CH₂ group is evidenced by a complex signal for H_{eq} at 2.47 ppm ($J_{gem} = 13.7, J_{vic} = 4.4$ Hz, W-coupling of 1.6 Hz) and by a d for H_{ax} at 1.98 ppm ($J_{gem} = 13.7$ Hz). Similar shifts and coupling constants were found for the orthoformate 5 [13]. Bu₃SnD allowed the introduction of a D label. Surprisingly, this deuteration was poorly selective, leading to a 55:45 mixture of DL-7 and DL-8 (97%). Acid hydrolysis of DL-6 gave selectively 2-O-pentanoyl-1,2,3,5/4-cyclohexanepentol



a) BuC(OMe)₃, camphorsulfonic acid, DMSO, 60°; 91%. b) 4-MeC₆H₄OC(= S)Cl, Et₃N, CH₂Cl₂, 23°; 91%. c) Bu₃SnH, AIBN, toluene, reflux; 80-93%. d) Conc. aq. HCl soln., MeOH, reflux; 100%. e) MeONa, MeOH, 23°; 85%. f) (t-Bu)Me₂SiOSO₂CF₃, 2,6-dimethylpyridine, CH₂Cl₂, 0 → 23°; 81%. g) (S)-Phenylethyl isocyanate, 4-(dimethylamino)pyridine (DMAP), CH₂Cl₂, reflux; 42% of D-12, 43% of L-13. h) LiBHEt₃, THF, 0 → 23°; 100% of D-11, 88% of L-11. i) TBAF · 3H₂O, THF, reflux; 57%. j) 4-BrC₆H₄COCl, DMAP, CH₂Cl₂, 23°; 44%. k) Conc. aq. HCl soln./MeOH, 23°; MeONa, MeOH, 23°; 78% of D-10, 75% of L-10.

(DL-9; > 99%). The position of the acyl group is readily deduced from the small coupling constant (2.6 Hz) of the strongly deshielded t at 6.06 ppm³). The pentanoate DL-9 was deacylated with MeONa in MeOH, whereupon analytically pure DL-10 precipitated in 85% yield. The ¹H-NMR spectrum and melting point of DL-10 are in agreement with literature data [5]. In this way, racemic 1,2,3,5/4-cyclohexanepentol (DL-10) was obtained from *myo*-inositol (1) in five steps and 65% overall yield, requiring one easy chromatography.

To prepare the enantiomerically pure 1,2,3,5/4-cyclohexanepentols D-10 and L-10, we silylated the diol DL-6 to yield 81% of the silyl ether DL-11. Treatment of DL-11 with (S)-phenylethyl isocyanate, followed by chromatography, gave the diastereoisomeric carbamates D-12 (42%) and L-13 (43%). These carbamates exist as mixture of rotamers, as indicated by the splitting of several peaks in the ¹³C-NMR spectra, and as already found for their orthoformate analogues D-14 and L-15 [13]. Reductive decarbamoylation of D-12 and L-13 with LiBHEt₃ gave the silylated orthopentanoates D-11 (100%) and L-11 (88%), respectively. The absolute configuration of D-11 was assigned on the basis of the positive first *Cotton* effect of the bis(4-bromobenzoate) D-16 [13][17], prepared from D-11 by desilylation and acylation. The required pentol D-10 was obtained in 78% yield by treating D-11 first with MeOH/HCl and then with MeOH/NaOMe. Similarly, L-11 yielded 75% of L-10. This synthesis provides D-10 and L-10 in eight steps from 1 and in a combined overall yield of 41-49%.

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Experimental Part

General. Solvents were freshly distilled from CaH₂ or Na/benzophenone. Anal. TLC: Merck precoated silica gel 60 F254 plates; detection by treatment with a soln. of 5% (NH₄)₆Mo₇O₂₆ · 4 H₂O, 0.1% Ce(SO₄)₂ · H₂O, in 10% H₂SO₄ soln. Flash chromatography (FC): silica gel Merck 60 (40–63 µm). High-performance liquid chromatography (HPLC): Spherisorb Silica (5 µm; prep. column: 250×20 mm; anal. column: 250×4 mm), UV detection (255 nm), $t_{\rm R}$ in min. M.p.: uncorrected. Optical rotations: 1-dm cell at 25° at 589 nm. UV Spectra: $\lambda_{\rm max}$ (ε) in nm. CD Spectra: λ ($\Delta\varepsilon$) in nm. FT-IR Spectra: absorption in cm⁻¹; conc. of the CCl₄ or CH₂Cl₂ soln. in mM. NMR Spectra: chemical shifts in ppm relative to TMS (¹H, ¹³C); coupling constants in Hz. Mass spectra: DCI at 70 eV; FAB in 3-nitrobenzyl alcohol (NBA) matrix.

1,3,5-O-Pentylidyne-myo-inositol (2). A suspension of myo-inositol (1) (8.34 g, 46.3 mmol) and camphorsulfonic acid (240.3 mg, 1.03 mmol) in DMSO (30 ml) was heated to 60°, treated with trimethyl orthopentanoate (8.55 ml, 48.6 mmol) over 2 h, and stirred for 15 h at 60°. The soln. was neutralized with Et₃N (280 µl, 2.00 mmol) and concentrated (75–79°, <1 Torr) until solidification of the residue. This residue was dissolved in hot AcOEt (60 ml) and filtered through a SiO₂ pad (elution with 500 ml of AcOEt). Evaporation afforded anal. pure **2** (11.40 g, 91%). White prisms. $R_f(Et_2O)$ 0.41. M.p. 127–128°. IR (sat. CCl₄ soln.): 3618m, 3584m, 3540m, 2961m, 1082s, 1056m. IR (4 mM, CH₂Cl₂): 3594m, 3577m, 3515m, 2962m, 1081s, 1057m. ¹H-NMR (300 MHz, CDCl₃): 4.56 (dt, J = 7.6, 3.8, H–C(4), H–C(6)); 4.26–4.22 (m, H–C(1), H–C(3), H–C(5)); 4.10 (dt, J = 11.8, 1.9, H–C(2)); 3.44 (br. d, J = 6.9, HO–C(4), HO–C(6)); 3.12 (d, J = 11.7, HO–C(2)); 1.74–1.69 (m, CH₂); 1.45–1.37 (m, CH₂); 1.31 (sext., J = 7.2, CH₂); 0.89 (t, J = 7.2, Me). ¹³C-NMR (50 MHz, CDCl₃): 108.99 (s, CO₃); 74.50 (d, C(1), C(3)); 68.14 (d, C(5)); 67.75 (d, C(4), C(6)); 59.56 (d, C(2)); 3.57 (t, CH₂); 24.45 (t, CH₂); 22.17 (t, CH₂); 1.3.62 (q, Me). ¹³C-NMR (75 MHz, CD₃OD): 110.26 (s, CO₃); 76.49 (d, C(1), C(3)); 70.60 (d, C(5)); 69.00 (d, C(4), C(6)); 50.43 (d, C(2)); 37.97 (t, CH₂); 25.82 (t, CH₂); 23.56 (t, CH₂); 14.31 (q, Me). DCI-MS (NH₄⁺): 331 (1, [M + BuCO]⁺), 247 (100, [M + 1]⁺), 115 (9), 102 (5), 85 (32, BuCO⁺). Anal. calc. for C₁₁H₁₈O₆ (246.26): C 53.65, H 7.37; found: C 53.47, H 7.25.

³) For an example in which the benzoyl group of a (partially protected) myo-inositol selectively migrates under acid catalysis from the equatorial O-C(3) to the axial O-C(2), see [16].

DL-1,3,5-O-Pentylidyne-4-O-[thioxo(tolyloxy)methyl]-myo-inositol (DL-4). a) A soln. of 2 (627.9 mg, 2.55 mmol) and Et₃N (888 µl, 6.37 mmol) in CH₂Cl₂ (25 ml) was treated with O-4-tolyl chlorothioformate for 11 min at 23° (caution: exothermic). Evaporation in vacuo (no heat) and treatment with Et₂O (30 ml) led to the precipitation of Et₃N·HCl. Filtration (fritted glass), evaporation, and chromatography (hexane/AcOEt 4:1 \rightarrow 1:1) gave 4 (915.6 mg, 91%). White needles.

b) Similarly, a soln. of **2** (2.873 g, 11.67 mmol) and Et₃N (4.07 ml, 29.16 mmol) in CH₂Cl₂ (50 ml) was treated with *O*-4-tolyl chlorothioformate (3.23 ml, 21.00 mmol) for 20 min at 23°, evaporated, taken in 30 ml of Et₂O, filtered through SiO₂, and evaporated. Crystallization from hexane/AcOEt 3:1 (60 ml) at -18° (15 h) yielded DL-4 (2.84 g, 61%). White needles. R_t (hexane/AcOEt 2:1) 0.51. M.p. 140–142°. IR (47.7 mM, CH₂Cl₂): 3599m, 3576m, 2963m, 2873w, 1506m, 1261s, 1219s, 1198s, 1080s, 1011m, 999m, 978m, 883m. ¹H-NMR (200 MHz, CDCl₃): 7.22 (d, J = 8.4, 2 arom. H); 6.97 (d, J = 8.4, 2 arom. H); 5.99 (td, J = 3.7, 1.7, H–C(4)); 4.59 (dtd, J = 7.8, 3.7, 1.6, H–C(6)); 4.54 (tt, J = 3.4, 1.7, H–C(5)); 4.47 (dq, J = 4.0, 2.0), 4.26 (dq, J = 4.0, 2.0, H–C(1), H–C(3)); 4.04 (dt, J = 12.0, D–C(2)); 3.05 (d, J = 12.0, HO–C(2)); 2.37 (s, Me); 2.33 (d, J = 7.8, HO–C(6)); 1.73–1.67 (m, CH₂); 1.51–1.41 (m, CH₂); 1.30 (sext., J = 7.3, CH₂); 0.89 (t, J = 3.7, 1.2, Hence C(6)); 1.73–1.67 (m, CH₂); 1.5103 (s, 1 arom. C); 136.71 (s, 1 arom. C); 130.14 (d, 2 arom. C); 121.16 (d, 2 arom. C); 109.86 (s, CO₃); 76.02 (d), 74.56 (d. C(1), C(3)); 71.35 (d, C(5)); 67.35 (d), 67.10 (d, C(4), C(6)); 60.08 (d, C(2)); 36.68 (t, CH₂); 24.56 (t, CH₂); 2.2.33 (t, CH₂); 20.87 (q, Me); 13.83 (q, Me). DCI-MS (NH⁺₄): 397 (2, $[M + 1]^+$), 365 (2, $[M - S + 1]^+$), 289 (100, $[M - Mec_6H_4O]^+$), 257 (19), 247 (13, $[M - Mec_6H_4OCS + 2]^+$), 108 (81, Mec₆H₄OH⁺). Anal. calc. for C₁₉H₂₄O₇S (396.46): C 57.56, H 6.10; found: C 57.58, H 5.92.

DL-1,3,5-O-Pentylidyne-1,2,3,5/4-cyclohexanepentol (DL-6). A soln. of DL-4 (1.69 g, 4.26 mmol) in toluene (40.0 ml) and a stock soln. of Bu₃SnH (2.00 ml, 7.53 mmol) and 2,2'-azobis(isobutyronitrile) (AIBN; 15.0 mg, 0.913 mmol) in toluene (15.0 ml) were degassed by bubbling Ar through them for 10 min. The soln. of DL-4 was heated to reflux, treated with the Bu₃SnH/AIBN soln. (first 10.0 ml, then 5.3 ml over 8 min), and stirred for 20 min. Evaporation and FC (hexane/AcOEt 2:1 \rightarrow 1:1) gave 6 (787.9 mg, 80%). Colourless oil. $R_{\rm f}$ (hexane/AcOEt 2:1) 0.25. IR (5 mM, CCl₄): 3628w, 3583w, 2961m, 2873w, 1558m, 1126m, 1082m, 1061m, 979m. IR (5 mM, CH₂Cl₂): 3603w, 3569w, 2961m, 1124m, 1083m, 1060m, 979m. 'H-NMR (200 MHz, CDCl₃): 4.53 (dt, J = 4.4, 3.7, D₂O shake \rightarrow t, J = 3.7, H–C(4)); 4.15–4.13 (m, H–C(1), H–C(3), H–C(5)); 3.81 (dt, J = 12.1, 2.1, H–C(2)); 3.03 (d, J = 12.1, HO–C(2)); 2.47 (dtd, J = 13.7, ca. 4.4, ca. 1.6, H_{eq}–C(6)); 1.98 (d, J = 13.7, H_{ax}–C(6)); 1.93 (d, J = 4.4, HO–C(4)); 1.67–1.62 (m, CH₂); 1.49–1.37 (m, CH₂); 1.30 (sext., J = 7.4, CH₂); 0.89 (t, J = 7.3, Me). ¹³C-NMR (75 MHz, CDCl₃): 110.04 (s, CO₃); 75.10 (d), 71.95 (d), 69.38 (d, C(1), C(3), C(5)); 65.80 (d, C(4)); 63.51 (d, C(2)); 37.77 (t, CH₂); 25.89 (t, C(6)); 24.44 (t, CH₂); 22.40 (t, CH₂); 1.80 (q, Me); DCI-MS (NH₄⁺): 315 (2, [M + BuCO]⁺), 231 (100, [M + 1]⁺), 99 (34), 85 (23, BuCO⁺).

DL-4-Deoxy-1,3,5-O-pentylidene-epi- $[4-{}^{2}H]$ inositol (DL-7) and DL-2-Deoxy-1,3,5-O-pentylidene-myo- $[4-{}^{2}H]$ inositol (DL-8). A soln. of DL-4 (103.6 mg, 0.261 mmol) in toluene (4.0 ml) and a soln. of Bu₃SnD (111 µl, 0.418 mmol) and AIBN (3.2 mg, 0.075 mmol) in toluene (0.9 ml) were degassed by bubbling Ar through them for 10 min. The soln. of DL-4 was heated to reflux, treated with the Bu₃SnD/AIBN soln. (first 0.4 ml, then the rest over 30 min), and stirred for 15 min. Evaporation and FC (hexane/AcOEt 2:1 \rightarrow 1:1) gave DL-7/DL-8 55:45 (58.9 mg, 97%). Colourless oil. ¹H-NMR (200 MHz, CDCl₃, 1 drop D₂O): 2.50-2.42 (m, 0.45 H, H_{eq}-C(4)); 1.95 (br. s, 0.55 H, H_{ax}-C(4)); other signals as for DL-6.

1DL-2-O-Pentanoyl-1,2,3,5/4-cyclohexanepentol (DL-9). A soln. of DL-6 (31.9 mg, 0.139 mmol) in MeOH (3.0 ml) and conc. aq. HCl (0.6 ml) was heated to reflux for 4 h. Evaporation gave DL-9 (34.2 mg, 100%, sole product according to ¹H-NMR). Colourless oil. $R_t(Et_2O) 0.00$. R_t (acetone) streaky 0.33–0.44. R_t (MeOH) 0.85. ¹H-NMR (300 MHz, (D₅)pyridine/CD₃OD 7:1: 6.06 (t, J = 2.6, H–C(2)); 4.27 (t, J = 9.2, H–C(4)); 4.19 (ddd, J = 11.1, 5.7, 2.8, irrad. at. 6.06 $\rightarrow dd$, J = 11.1, 5.7, irrad. at 2.51 $\rightarrow d$, J = 2.8, H–C(1)); 4.04–3.95 (m, H–C(5)); 3.99 (dd, J = 9.6, 2.8, H–C(3)); 2.53–2.46 (m, 2 H–C(6)); 2.30 (t, J = 7.5, CH₂); 1.53 (quint., J = 7.5, CH₂); 1.18 (sext., J = 7.5, CH₂); 0.64 (t, J = 7.3, Me). ¹³C-NMR (75 MHz, CDCl₃): 175.31 (s, C=O); 76.55 (d), 76.16 (d), 72.22 (d), 70.94 (d, C(1), C(2), C(3), C(5)); 66.90 (d, C(4)); 36.84 (t, CH₂); 35.06 (t, C(6)); 28.17 (t, CH₂); 23.22 (t, CH₂); 14.11 (q, Me).

1DL-1,3,5-O-Pentylidyne-1,2,3,5/4-cyclohexanepentol (DL-10). A soln. of DL-6 (112.5 mg, 0.48 mmol) in MeOH (3 ml) and sat. aq. HCl soln. (3 ml) was heated to reflux for 4 h and evaporated. The residue was dried azeotropically with EtOH (3 × 20 ml) dissolved in MeOH (11 ml), and treated with a soln. of MeONa (0.86M in MeOH, 7.3 mmol) in MeOH for 15 h at 23°. The precipitate was collected by filtration and washed with MeOH to give DL-10 (40.3 mg, 50%) as a white powder. The combined MeOH solns. were concentrated to 2 ml to give additional DL-10 (27.2 mg, 35%) as a white precipitate. White powder. R_f (acetone) 0.00. R_f (MeOH) streaky 0.00–0.23. M.p. 218–219° ([5]: > 220°, [10]: 208–210°). IR (KBr): 3404s, 3289s, 3167s, 2952w, 2874w, 1453m, 1377w, 1169w, 1071m, 993w, 918w, 816w, 722w, 662w, 600w. ¹H-NMR (300 MHz, D₂O): 3.99 (br. s, H--C(2));

3.78 (*ddd*, J = 12.1, *ca*. 3.7, *ca*. 2.8, H–C(1)); 3.55–3.46 (*m*, H–C(4), H–C(5)); 3.42 (*dd*, J = 10.0, 2.8, H–C(3)); 1.97 (*dt*, J = 11.8, 3.7, H_{eq}–C(6)); 1.75 (*q*, J = 11.8, H_{ax}–C(6)). ¹³C-NMR (75 MHz, D₂O): 77.14 (*d*); 75.67 (*d*); 74.60 (*d*); 72.11 (*d*); 69.32 (*d*); 36.60 (*t*). ESI-MS: 351 (30), 348 (22), 266 (22), 187 (59, [*M* + Na]⁺), 182 (100, [*M* + NH₄]⁺), 165 ([*M* + 1]⁺). Anal. calc. for C₆H₁₂O₅ (164.16): C 43.90, H 7.37; found: C 43.77, H 7.77.

IDL-2-O-[(tert-Butyl)dimethylsily]]-1,3,5-O-pentylidyne-1,2,3,5/4-cyclohexanepentol (DL-11). A soln. of DL-6 (372.3 mg, 1.62 mmol) and 2,6-dimethylpyridine (340 µl, 2.92 mmol) in CH₂Cl₂ (5.0 ml) was treated with (t-Bu)Me₂SiOSO₂CF₃ (400 µl, 1.74 mmol), stirred for 5 h at 23°, and poured into sat. aq. NaHCO₃ soln. Extraction with AcOEt, washing (brine), drying (Na₂SO₄), evaporation, and FC (hexane/AcOEt 1:8 \rightarrow 2:1) gave DL-11 (451.0 mg, 81%). Colourless oil. R_t (hexane/AcOEt 4:1) 0.25. R_t (hexane/CH₂Cl₂/AcOEt 6:2:1) 0.39. Anal. HPLC: t_R (hexane/AcOEt 16:1, 2 ml/min) 25.9. IR (17 mM, CCl₄): 3628w, 2957m, 2858m, 1471w, 1390m, 1251m, 1142m, 1120m, 1071m, 979m, 887m. IR (17 mM, CH₂Cl₂): 3605w, 2960m, 2858m, 1471w, 1389m, 1139m, 1117m, 1078m, 1071m, 979m, 887m. ¹H-NMR (200 MHz, CDCl₃): 4.54–4.44 (m, D₂O shake \rightarrow br. $t, J \approx 4.0, H-C(4)$); 4.50 (td, J = 3.7, 1.7, H-C(5)); 4.10–4.06 (m, H-C(1), H-C(3)); 3.93 (t, J = 1.9, H-C(2)); 2.43 ($dg, J = 13.3, ca. 2, H_{eq}-C(6)$); 2.29 (d, J = 4.6, OH); 1.93 (br. $d, J = 13.3, H_{ax}-C(6)$); 1.71–1.63 (m, CH_2); 1.56–1.32 (m, CH_2); 1.32 (sext., $J = 7.0, CH_2$); 0.95 (s, t-BuSi); 0.89 (t, J = 7.4, Me); 0.12 (s, Me_2 Si). ¹³C-NMR (75 MHz, CDCl₃): 109.79 (s, CO_3); 75.48 (d), 72.02 (d), 69.89 (d, C(1), C(3), C(5)); 66.48 (d, C(4)); 63.78 (d, C(2)); 37.80 (t, CH_2); 26.54 (t, C(6)); 25.73 (q, Me_3C); 24.55 (t, CH_2); 22.43 (t, CH_2); 18.13 (s, Me_3C); 13.84 (q, Me); -4.76 (q, Me_2 Si). DCI-MS (NH₄⁺): 345 (100, [M + 1]⁺), 287 (31, [$M - {}^{H}Bu$]⁺), 213 (13, [$M - {}^{H}BuMe_2$ SiO]⁺), 185 (49).

Treatment of DL-11 with (S)-Phenylethyl Isocyanate. A soln. of DL-11 (1.17 g, 3.48 mmol), sublimed DMAP (1.43 mg, 11.7 mmol), and (S)-phenylethyl isocyanate (717 μ l, 5.09 mmol) in CH₂Cl₂ (10.0 ml) was heated to reflux for 13 h, treated with additional (S)-phenylethyl isocyanate (300 μ l, 2.13 mmol), and stirred for 3 h. The excess isocyanate was hydrolysed with sat. aq. NaHCO₃ soln. (2 ml) at reflux for 30 min, wherupon a white precipitate appeared. The mixture was diluted with Et₂O (100 ml), filtered, washed with H₂O, aq. CuSO₄ soln., H₂O, and brine, dried (Na₂SO₄), and evaporated. FC (hexane/CH₂Cl₂/AcOEt 6:2:1) gave D-12/L-13 (1.58 g, 95%) which were separated by prep. HPLC (hexane/CH₂Cl₂/AcOEt 70:21:9): D-12 (706.3 mg, 42%) and L-13 (717.6 mg, 43%). Colourless oils.

 $1D-2-O-[/(tert-Butyl) dimethylsily] - 1,3,5-O-pentylidyne-4-O-{[/(S)-1-phenylethyl]carbamoyt]-1,2,3,5/4-cy-clohexanepentol (D-12): R₁(hexane/CH₂Cl₂/AcOEt 6:2:1) 0.58. Prep. HPLC: t_R (hexane/CH₂Cl₂/AcOEt 70:21:9, 10 ml/min) 14.0 min. [z]_D²⁵ = -12.8 (c = 3.17, CCl₄). IR (65 mM, CCl₄): 3450w, 3275w, 2960s, 2858m, 1737s, 1495m, 1389m, 1250m, 1216m, 1144s, 1123m, 1072m, 981m, 886m. ¹H-NMR (300 MHz, 50°, CDCl₃): 7.37-7.25 (m, 5 arom. H); 5.29-5.23 (br. s), 5.04-4.86 (br. s, NH, H-C(4)); 4.97-4.65 (br. s, PhCH); 4.24-4.15 (br. s, 2 H), 4.12-3.95 (br. s), 3.83-3.60 (br. s, H-C(1), H-C(2), H-C(3), H-C(5)); 2.43 (br. d, J = 13.4, H_{eq}-C(6)); 1.73-1.62 (m, H_{ax}-C(6)); 1.66-1.61 (m, CH₂); 1.49 (d, J = 6.9, Me); 1.48-1.40 (m, CH₂); 1.31 (sext., J = 7.3, CH₂); 0.93 (s, t-Bu); 0.87 (t, J = 7.3, Me); 0.08 (s, Me₂Si). ¹³C-NMR (75 MHz, 50°, CDCl₃): 75.40 (br. s, CC); 12.587 (br. d, 1 arom. C); 129.80 (d, 2 arom. C); 127.57 (d, 2 arom. C); 125.87 (br. d, 1 arom. C); 109.99 (s, CO₃); 72.92 (d, 0.5 C), 72.79 (d, 0.5 C), 71.50 (d, 0.5 C), 71.34 (d, 0.5 C, (3), (C5)); 68.73 (br. d, C(1)); 67.47 (d, 0.5 C), 67.33 (d, 0.5 C, C(4)); 64.20 (br. d, C(2)); 50.99 (br. d, CHN); 37.61 (t, CH₂); 27.25 (br. t, C(6)); 25.60 (q, Me₃C); 24.42 (t, CH₂); 22.30 (t, CH₂); 17.98 (s, Me₃C); 13.68 (q, Me); -4.84 (br. q, Me₂Si). FAB-MS (NOBA): 925 (1, [2M - ¹Bu]⁺), 587 (3, [M + 96]⁺), 492 (90, [M + 1]⁺), 476 (11, [M - Me]⁺), 434 (100, [M - ¹Bu]⁺), 327 (25, [M - PhMeCHNHCOO]⁺). Anal. calc. for C₂₆H₄₁NO₆Si (491.70): C 63.51, H 8.40, N 2.85; found: C 63.24, H 8.14, N 2.84.$

1L-2-O-[(tert-Butyl)dimethylsily]-1,3,5-O-pentylidyne-4-O-{[(S)-1-phenylethyl]carbamoyl}-1,2,3,5/4-cyclohexanepentol (L-13): $R_{\rm f}$ (hexane/CH₂Cl₂/AcOEt 6:2:1) 0.49. Prep. HPLC: $t_{\rm g}$ (hexane/CH₂Cl₂/AcOEt 70:21:9, 10 ml/min) 18.0 min. [a]_D²⁵ = -42.0 (c = 2.58, CCl₄). IR (52 mM, CCl₄): 3450w; 3276w; 2960s, 2858m, 1737s, 1721s, 1495m, 1389m, 1250m, 1217m, 1144s, 1123m, 1072m, 981m, 886m. ¹H-NMR (300 MHz, 50°, CDCl₃): 7.37-7.27 (m, 5 arom. H); 5.31-5.23 (br. s), 5.10-4.80 (br. s, NH, H-C(4)); 4.97-4.69 (br. s, PhCH); 4.30-4.05 (br. s), 4.23-4.08 (br. s), 4.11-3.92 (br. s), 3.78-3.55 (br. s, H-C(1), H-C(2), H-C(3), H-C(5)); 2.57-2.32 (br. s), 8.10, 2.35-1.97 (br. s, 0.2 H, H_{eq}-C(4)); 1.82-1.64 (br. s, H_{ax}-C(4)); 1.65-1.60 (m, CH₂); 1.53-1.39 (m, 5 H, CH₂, Me); 1.30 (sext., J = 7.3, CH₂); 0.92 (s, t-Bu); 0.87 (t, J = 7.3, Me); 0.08 (s, Me₂Si). ¹³C-NMR (75 MHz, 50°, CDCl₃): 154.04 (br. s, C=O); 143.05 (br. s, 1 arom. C); 128.64 (d, 2 arom. C); 127.37 (d, 1 arom. C); 125.76 (br. d, 2 arom. C); 109.85 (s, CO₃); 72.84 (d), 71.31 (d), 68.56 (d, C(1), C(3)); 67.22 (br. d, C(4); 64.21 (d, C(2)); 52.08 (br. d, 0.2 C), 50.56 (br. d, 0.8 C, CH-N); 37.41 (t, CH₂); 27.19 (br. t, 0.8 C, C(6)); 25.54 (q, Me_3 C); 24.34 (t, CH₂); 22.24 (t, CH₂); 21.76 (br. t, 0.2 C, C(6)); 17.90 (s, Me₃C); 13.64 (q, Me); -4.83 (q, MeSi); -4.93 (q, MeSi). FAB-MS (NOBA): 983 (1, [2M + 1]⁺), 925 (2, [2M -'Bu]⁺), 587 (7, [M + 96]⁺), 492 (100, [M + 1]⁺), 476 (10, [M -Me]⁺), 434 (82, [M -'Bu]⁺), 327 (17, [M - PhMe-CHNHCOO]⁺). Anal. calc. for C₂₆H₄₁NO₆Si (491.70): C 63.51, H 8.40, N 2.85; found: C 63.29, H 8.18, N 2.88. 1D-2-O-[(tert-Butyl)dimethylsilyl]-1,3,5-O-pentylidyne1,2,3,5/4-cyclohexanepentol (D-11). A soln. of D-12 (255.7 mg, 0.520 mmol) in THF (5.0 ml) was cooled to 0° and treated dropwise with a soln. of 1.0M LiBHEt₃ (2.08 mmol) in THF over 3 min (*caution*: fizz). The soln. was stirred at 23° for 3 h, diluted with i-Pr₂O (10 ml), cooled to 0°, and treated with a. phosphate buffer (pH 7.0, 1.0M, 6 ml; *caution*: fizz) and by 30% aq. H₂O₂ soln. (0.5 ml; *caution*: highly exothermic, fizz). After 10 min of vigorous stirring at 23°, extraction (AcOEt), washing (H₂O, brine), drying (Na₂SO₄), and FC (hexane/(CH₂Cl₂/AcOEt 2:1) 4:1 \rightarrow 0.7:1) afforded D-11 (178.2 mg, 100%) as a colourless oil, which crystallized from hexane at -18°. White plates. M.p. 80-81°. [α]_D²⁵ = + 3.6 (c = 4.38, CCl₄). Anal. calc. for C₁₇H₃₂O₅Si (344.52): C 59.27, H 9.36; found: C 59.31, H 7.33.

1L-2-O-[(tert-Butyl)dimethylsilyl]-1,3,5-O-pentylidyne-1,2,3,5/4-cyclohexanepentol (L-11). The equivalent procedure applied to L-13 (636.4 mg, 1.29 mmol) afforded L-11 (392 mg, 88%).

1D-1,3,5-O-Pentylidyne-1,2,3,5/4-cyclohexanepentol (D-6). A soln. of D-11 (87.1 mg, 0.253 mmol) and Bu₄NF · 3H₂O (240 mg, 0.76 mmol) in THF (4.4 ml) was heated to reflux for 2 h. The soln. was diluted with AcOEt, washed with H₂O and brine, dried (Na₂SO₄), and evaporated. FC (hexane/AcOEt 4:1 \rightarrow 2:1) afforded D-6 (33.0 mg, 57%) as a colourless oil, which crystallized from pentane/ⁱPr₂O. White plates. M.p. 129–130°. [α]_D²⁵ = + 1.3 (c = 1.31, CHCl₃). Anal. calc. for C₁₁H₁₈O₅ (230.26): C 57.38, H 7.88; found: C 57.35, H 7.67.

1D-2,4-Bis-O-(4-bromobenzoyl)-1,3,5-O-pentylidyne-1,2,3,5/4-cyclohexanepentol (D-16). A soln. of D-6 (13.1 mg, 56.9 μmol), DMAP (114.2 mg, 936 μmol), and 4-bromobenzoyl chloride (155.0 mg, 706 μmol) in CH₂Cl₂ (1.0 ml) was stirred for 15 h at 23°. The resulting suspension was diluted with Et₂O, washed (H₂O, aq. CuSO₄ soln., brine), dried (Na₂SO₄), and evaporated. FC (hexane/ACOEt 16:1 → 8:1) afforded D-16 (15.0 mg, 44%). Colourless oil. R_f (hexane/ACOEt 9:1) 0.46. UV (20 μM, MeCN): 246 (48200). CD (20 μM, MeCN): 241 (0), 251 (+ 16.5). ¹H-NMR (200 MHz, CDCl₃): 7.99 (d, J = 8.7, 2 arom. H); 7.88 (d, J = 8.4, 2 arom. H); 7.63 (d, J = 8.7, 2 arom. H); 7.61 (d, J = 8.4, 2 arom. H); 5.73 (td, J = 4.4, 1.9, H-C(4)); 5.18 (t, J = 1.9, H-C(2)); 4.58 (dq, J = 4.0, 2.0, H-C(3)); 4.50-4.44 (m, H-C(1), H-C(5)); 2.68 (dtd, J = 14.0, 4.0, 1.6, H_{eq}-C(6)); 2.04 (dt, J = 14.0, ca. 1.0, H_{ax}-C(6)); 1.77-1.72 (m, CH₂); 1.53-1.46 (m, CH₂); 1.36 (sext., J = 7.4, CH₂); 0.89 (t, J = 7.3, Me).

1L-1,2,3,5/4-Cyclohexanepentol (L-10). A soln. of L-11 (161.5 mg, 0.47 mmol) in MeOH (2 ml) and conc. HCl soln. (2 ml) was stirred for 45 min at 23° and evaporated. The residue was dried azeotropically with EtOH (3 × 15 ml), dissolved in MeOH (5 ml), and treated with a soln. of MeONa (0.86M in MeOH, 0.09 mmol) in MeOH for 8 h at 23°. The precipitate was collected by filtration and washed with MeOH to give L-10 (50.8 mg, 66%) as a white powder. The combined MeOH solns. were concentrated to 2 ml to give additional D-10 (9.8 mg, 12%) as a white precipitate. M.p. 195-196°. $[\alpha]_D^{25} = - 8.0$ (c = 0.64, H₂O).

1D-1,2,3,5/4-cyclohexanepentol (D-10). The equivalent procedure applied to D-11 (98.2 mg) afforded D-10 (35.2 mg). M.p. 194-195°. $[\alpha]_{D^5}^{25} = +8.9 (c = 0.5, H_2O).$

REFERENCES

- R. Irvine, P. Cullen, Curr. Biol. 1996, 6, 537; D. C. Billington, Chem. Soc. Rev. 1989, 18, 83; B. V. L. Potter, S. R. Nahorski, Biochem. Soc. Trans. 1993, 20, 434; N. Divecha, R. F. Irvine, Cell 1995, 80, 269.
- [2] A. Crossman, Jr., J. S. Brimacombe, M. A. J. Ferguson, J. Chem. Soc., Perkin Trans. 1 1997, 2769;
 P. T. Englund, Ann. Rev. Biochem. 1993, 62, 121; M. A. J. Ferguson, Biochem. Soc. Trans. 1993, 20, 243.
- [3] T. Suami, S. Ogawa, Y. Funaki, Bull. Chem. Soc. Jpn. 1975, 48, 1545.
- [4] C. Jiang, J. D. Moyer, D. C. Baker, J. Carbohydr. Chem. 1987, 6, 319.
- [5] S. V. Ley, M. Parra, A. J. Redgrave, F. Sternfeld, Tetrahedron 1990, 46, 4995.
- [6] J. Cleophax, D. Dubreuil, S. D. Gero, A. Loupy, M. Vieira de Almeida, A. D. da Silva, G. Vass, Bioorg. Med. Chem. Lett. 1995, 5, 831.
- [7] F. McPhee, P. Downes, G. Lowe, Biochem. J. 1991, 277, 407.
- [8] K. M. Pietrusiewicz, G. M. Salamonczyk, Synth. Commun. 1995, 25, 1863.
- [9] S. Ballereau, P. Guédat, B. Spiess, N. Rehnberg, G. Schlewer, Tetrahedron Lett. 1995, 36, 7449; J. L. Offer, H. P. Voorheis, J. C. Metcalfe, G. A. Smith, J. Chem. Soc., Perkin Trans. 1 1992, 953.
- [10] J. D. Moyer, O. Reizes, S. Ahir, C. Jiang, N. Malinkowski, D. C. Baker, Mol. Pharmacol. 1988, 33, 683.
- [11] S. C. Cosulich, J. Offer, G. A. Smith, R. Hesketh, J. C. Metcalfe, Biochem. J. 1993, 292, 719.
- [12] M. A. Biamonte, A. Vasella, Helv. Chim. Acta 1998, 81, 695.
- [13] A. Zapata, B. Bernet, A. Vasella, Helv. Chim. Acta 1996, 79, 1169.
- [14] H. W. Lee, Y. Kishi, J. Org. Chem. 1985, 50, 4402; G. Baudin, B. I. Glänzer, K. S. Swaminathan, A. Vasella, Helv. Chim. Acta 1988, 71, 1367.

- [15] P. Uhlmann, A. Vasella, Helv. Chim. Acta. 1992, 75, 1979.
- [16] J. L. Meek, F. Davidson, J. F. W. Hobbs, J. Am. Chem. Soc. 1988, 110, 2317.
- [17] M. Chang, H. V. Meyers, K. Nakanishi, M. Ojita, J. H. Parker, M. H. Parker, R. Takeda, J. T. Vázquez, W. T. Wiesler, Pure Appl. Chem. 1989, 61, 1193.

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